

This article was downloaded by:

On: 26 January 2011

Access details: *Access Details: Free Access*

Publisher *Taylor & Francis*

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Nucleosides, Nucleotides and Nucleic Acids

Publication details, including instructions for authors and subscription information:

<http://www.informaworld.com/smpp/title~content=t713597286>

Piperidinyl Peptide Nucleic Acids: Synthesis and DNA-Complementation Studies

Pallavi Lonkar^a; Vaijayanti A. Kumar^a

^a Division of Organic Chemistry, National Chemical Laboratory, Pune, India

Online publication date: 09 August 2003

To cite this Article Lonkar, Pallavi and Kumar, Vaijayanti A.(2003) 'Piperidinyl Peptide Nucleic Acids: Synthesis and DNA-Complementation Studies', *Nucleosides, Nucleotides and Nucleic Acids*, 22: 5, 1105 — 1108

To link to this Article: DOI: 10.1081/NCN-120022747

URL: <http://dx.doi.org/10.1081/NCN-120022747>

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: <http://www.informaworld.com/terms-and-conditions-of-access.pdf>

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

Piperidinyl Peptide Nucleic Acids: Synthesis and DNA-Complementation Studies

Pallavi Lonkar and Vijayanti A. Kumar*

Division of Organic Chemistry, National Chemical Laboratory,
Pune, India

ABSTRACT

Synthesis of a new six membered PNA analogue by introducing a methylene bridge between β carbon atom of ethylene diamine and β' carbon atom of linker to nucleobase.

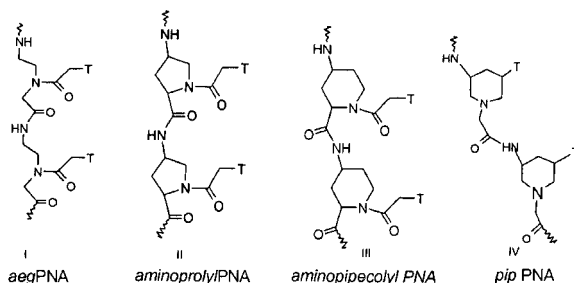
INTRODUCTION

Peptide Nucleic Acids have emerged as potential antisense agents.^[1] They bind to DNA in both parallel as well as antiparallel orientation. To discriminate between this binding property one can introduce chirality in PNA. In earlier work from our laboratory a built-in chirality was achieved by introducing a methylene bridge^[2] between the α -C atom of the glycyl unit and the β -C atom of the ethylene segment of *aeg*PNA(I) which gave *pr*PNA(II). The homooligomers derived from these *aminopropyl* PNA monomer do not bind to the complementary DNA sequence,

*Correspondence: Vijayanti A. Kumar, Division of Organic Chemistry, National Chemical Laboratory, Synthesis, Dr. Homi Bhabha Road, Pune 411008, India; Fax: +91 20 5893355; E-mail: vkumar@dna.ncl.res.in.



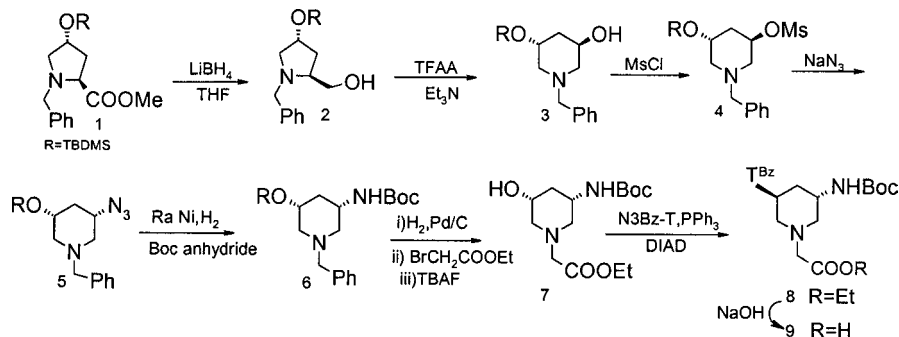
probably due to conformational rigidity in the oligomer. To release this conformational rigidity, aminoethyl backbone in *aminopropyl* PNA was replaced by amino-propyl backbone still retaining the two chiral centers to obtain *pipecolyl*/PNA (III)^[3] which too was found to significantly impair the complexation of PNA with complementary DNA.



Herein, we report synthesis of a new six membered PNA analogue by introducing a methylene bridge between β carbon atom of ethylene diamine and β' carbon atom of linker to nucleobase. (*pip* PNA molecules with a methylene bridge inserted between α carbon atom of ethylenediamine and β' carbon atom of linker to nucleobase and simultaneously removing rigid carbonyl group. Their DNA/RNA binding preferences may be dictated by the geometry of the backbone as well as the orientation of the nucleobase.

RESULTS AND DISCUSSION

The suitably protected *trans*-4-hydroxy proline **1** was converted to the *trans*-2*S*,4*R* pyrrolidine-2-methanol **2** by reduction of ester function. Treatment with tri-fluoroacetic anhydride followed by triethylamine gives the six membered rearranged product **3** with retention of configuration. Mesylation of the resulting unprotected hydroxy group and reaction with sodium azide gave **5** with inversion at C3. Compound **5** was then selectively hydrogenated using Ra-Ni and Boc-protected to get amino piperidine derivative **6** as shown in Scheme 1. Compound **6** was then



Scheme 1.

Table 1. UV Tm studies of PNA2:DNA complexes.

	Sequences	UV Tm°C
PNA10	H-TTTTTTTT-β-ala-OH	43
PNA11	H-TTTTTTTt-β-ala-OH	50.7
PNA12	H-TTTtTTTT-β-ala-OH	54.4
PNA13	H-TTTtTTTT-β-ala-OH	41.4
PNA14	H-tTTTTTTT-β-ala-OH	—
DNA	5'-GC(A) ₁₀ CG-3'	

t indicates modified PNA unit.

subjected to hydrogenation, alkylation of ring nitrogen using ethylbromoacetate followed by removal of silyl protection using TBAF to get **7**. *Trans*-5*S*-N3-benzoyl-Thymin-1-yl-3*S*-Bocaminomethyl pyrrolidine derivative **8** was synthesized under Mitsunobu conditions. This was then hydrolyzed using aqueous methanolic sodium hydroxide to get the thymine monomer **9** that could be used for solid phase synthesis of PNA-PyrrolidinePNA oligomer/mixmers. All the new compounds were characterized using suitable spectroscopic analysis.

PNA oligomers containing the *aeg*PNA and piperidine-PNA backbone units were synthesized by SPPS using the BOC- protection strategy. DNA oligomers were synthesized on Pharmacia GA plus synthesizer employing phosphoramidite chemistry.

The PNA10 is the unmodified PNA10 with aminoethylglycyl backbone. PNA11-PNA14 are the modified PNA oligomers with the modified PNA units incorporated at the predefined sites as represented in Table 1. The UV-Tm studies of these monomers indicate that the modified PNA unit at the C-terminus in PNA 11 stabilizes the complex with complementary DNA by about 7°C. The synergistic effect is observed with one more unit in the center of the sequence PNA12 as the PNA₂:DNA complex is further stabilized by about 47°C. Modified unit only in the center of the sequence PNA13 causes 2°C destabilization. These preliminary results are very encouraging and need to be further investigated.

CONCLUSIONS

A high yielding stereospecific ring expansion of protected hydroxy prolinol gives suitably substituted piperidine ring. The ring nitrogen is protonated at physiological pH and oligomers are highly water-soluble. DNA complementation studies by UV-Tm measurements indicate that the six membered monomer is capable of stabilizing the PNA₂:DNA complexes.

ACKNOWLEDGMENTS

VAK thanks Department of Science and Technology, New Delhi, and National Chemical Laboratory, Pune, for financial support. Meena thanks CSIR, New Delhi, for research fellowship.



REFERENCES

1. Nielsen, P.E.; Egholm, M.; Berg, R.H.; Buchardt, O. *Science* **1991**, *254*, 1497.
2. Gangamani, B.P.; Kumar, V.A.; Ganesh, K.N. *Tetrahedron* **1999**, *55*, 177.
3. Lonkar, P.S.; Kumar, V.A.; Ganesh, K.N. *Nucleosides Nucleotides Nucleic Acids* **2001**, *20*, 1197.
4. Kumar, V.A.; Pallan, P.S.; Meena; Ganesh, K.N. *Org. Lett.* **2001**, *3*, 1269–1272.

